

## HIV1 RNA quantification

### History

Developmental work 1997 to 1998

Introduced 1999

Accredited to ISO 17043

### Review

Ten Years of External Quality Assessment of Human Immunodeficiency Virus Type 1 RNA Quantification

Senechal and James

J. Clin. Micro. 2012: 50 (11), 3614-3619



## Scheme format

- Lyophilised specimens
- Normal human plasma diluent
- Paired specimens prepared from a single HIV RNA positive plasma to represent sequential specimens from an individual patient
- Data analysis based on the difference in concentration between the specimens

# Results presentation and performance evaluation

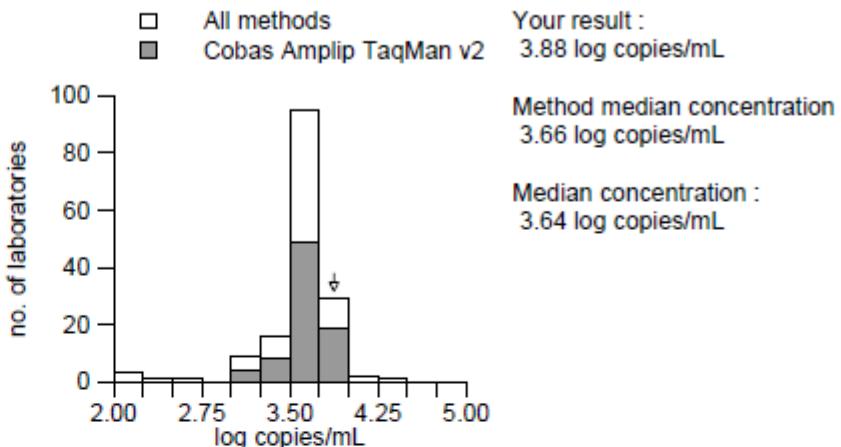
- No reference standard available when the scheme was introduced
- Absolute values
  - Consensus result
    - Promotes comparability and is an ideal objective
    - Over 12% failure to report to within 0.5 log copies of the median (2010 figures);  
~7% (2014 figures)
  - Consensus by kit
    - Use of in-house assays would exclude evaluation for some participants
- Difference in viral load between paired specimens
  - Clinically relevant
  - Avoids bias due to differences in absolute values obtained with different manufacturers' kits
  - Not affected by differences in reporting units

# Report excerpt: distribution 3480, June 2014

Specimens 2061 and 2062 originated from a serial dilution from a single HIV-1 RNA plasma (subtype C) diluted 1:2.9 and 1:16 respectively in negative human plasma, yielding a theoretical difference of 0.74 log copies/mL.

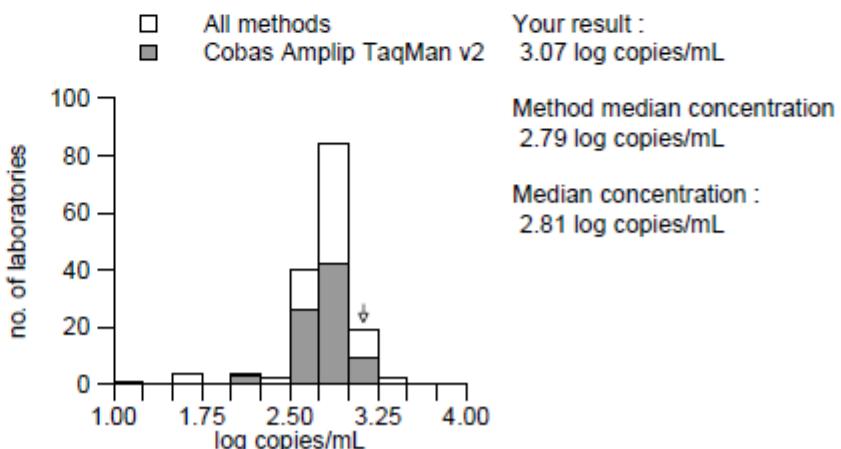
## Specimen : 2061

	n (UK)	range	median	5%-95%
All methods	164 (54)	2.00-7.00	3.64	3.11-3.90
Abbott Real-Time	37 (17)	3.38-4.17	3.65	3.46-3.89
Cobas Amplip TaqMan v2	80 (23)	3.03-3.92	3.66	3.29-3.88
Cobas HP TaqMan v2	10 (1)	3.11-7.00	3.59	3.11-5.56
Nuclisens Easy-Q v2	6 (3)	2.00-2.62	2.27	2.05-2.59
Qiagen: Artus	8 (4)	3.56-4.23	3.77	3.59-4.19
RAS Amplisure	1	3.24-3.24		
Real-Time Single target	2 (2)	3.83-4.40		
Roche: Cobas TaqMan	7 (2)	3.03-3.79	3.56	3.17-3.77
Siemens: kPCR	8	3.56-3.69	3.63	3.57-3.68
Unspecified	1 (1)	3.42-3.42		



## Specimen : 2062

	n (UK)	range	median	5%-95%
All methods	161 (53)	1.00-3.73	2.81	2.25-3.08
Abbott Real-Time	37 (17)	2.64-3.38	2.90	2.72-3.12
Cobas Amplip TaqMan v2	80 (23)	2.15-3.12	2.79	2.59-3.07
Cobas HP TaqMan v2	10 (1)	1.00-3.01	2.72	1.63-2.99
Nuclisens Easy-Q v2	4 (2)	1.54-1.71	1.63	1.56-1.70
Qiagen: Artus	8 (4)	2.73-3.73	2.98	2.75-3.52
Real-Time Single target	2 (2)	2.78-3.30		
Roche: Cobas TaqMan	7 (2)	2.15-3.05	2.74	2.29-3.01
Siemens: kPCR	8	2.61-2.89	2.85	2.63-2.88
Unspecified	1 (1)	2.67-2.67		



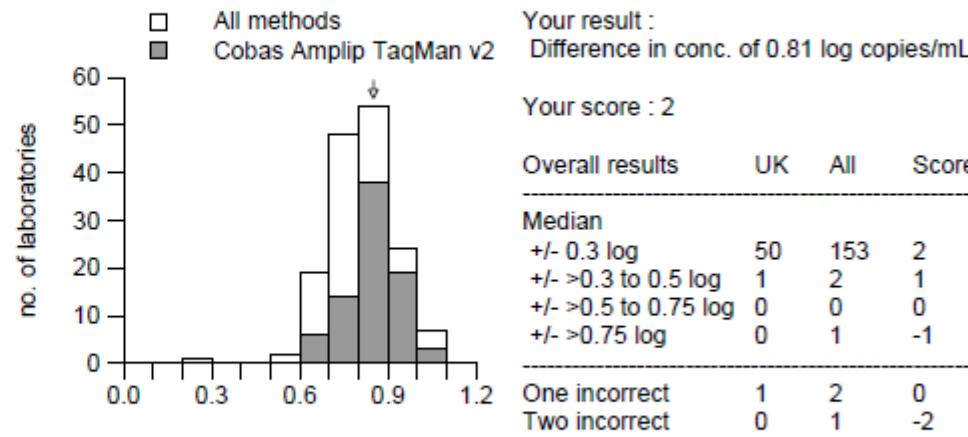
# Report excerpt: distribution 3480, June 2014

Intended result : 0.48 to 1.08 log copies/mL

(average median of the reported differences in concentration between specimen 2061 and 2062 +/- 0.3 log copies/mL, with an uncertainty of 0.043)

Difference in concentration between specimen 2061 and 2062 expressed in log copies/mL :

	n (UK)	range	av.median	5%-95%
All methods	156 (51)	0.29-6.00	0.78	0.65-1.00
Abbott Real-Time	36 (16)	0.57-0.99	0.73	0.66-0.85
Cobas Amplip TaqMan v2	80 (23)	0.63-1.02	0.85	0.68-0.97
Cobas HP TaqMan v2	10 (1)	0.57-6.00	0.74	0.62-3.71
Nuclisens Easy-Q v2	4 (2)	0.29-0.98	0.67	0.35-0.93
Qiagen: Artus	7 (4)	0.64-1.00	0.85	0.66-0.98
Real-Time Single target	2 (2)	1.05-1.10		
Roche: Cobas TaqMan	7 (2)	0.74-0.91	0.82	0.76-0.90
Siemens: kPCR	8	0.75-1.04	0.80	0.75-0.98
Unspecified	1 (1)	0.75-0.75		



A total of 179 sets of specimens were distributed for testing with 163 participants responding within the specified period: four participants did not examine the specimens. Valid results for both specimens were reported by 156 participants with 153/156 (98%) reporting results within 0.30 log copies/mL of the average of the median difference in concentration.

Of the outlying results: two participants reported a result within 0.3 to 0.5 log copies/mL (Method: Nuclisens Easy-Q v2, Real-Time Single target) and one participant reported a result over 0.75 log copies/mL (Method: Cobas High Pure TaqMan v2). Two participants reported a false negative result for specimen 2062 (Method: Unspecified and Nuclisens Easy-Q v2), this is categorised as 'One incorrect'. One participant reported a false negative result for both specimens (Method: Sacace Real-TM).

# Contact details

Dr Vivienne James

**UK NEQAS for Microbiology**

PO Box 63003

London NW9 1GH

Tel: + 44 (0)20 8905 9890

Fax: + 44 (0)20 8205 1488

Email: [organiser@ukneqasmicro.org.uk](mailto:organiser@ukneqasmicro.org.uk)

Website: [www.ukneqasmicro.org.uk](http://www.ukneqasmicro.org.uk)